

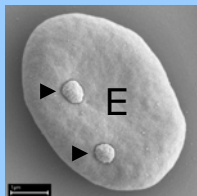
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Introduction

Hemotropic mycoplasmas cause infectious anemia in several species (1).



Erythrocyte with two coccoid bodies of *Mycoplasma haemofelis*

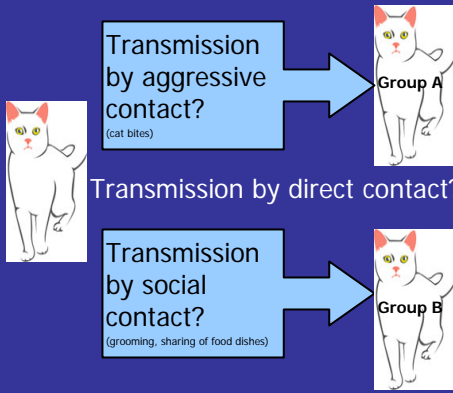
In cats three hemotropic mycoplasmas (aka hemoplasmas) are known: *Mycoplasma haemofelis*, *Candidatus Mycoplasma haemominutum* and *Candidatus Mycoplasma turicensis* (2,3).

The natural transmission routes of these agents are largely unknown (4).

Saliva samples of infected cats were found hemoplasma PCR-positive, but loads were low (5).

Aim of the study

The aim of the study was to evaluate possible routes of transmission of *Candidatus Mycoplasma turicensis* via direct contact among cats and to confirm successful infections by qPCR and serodiagnostics.



Material & Methods

For the transmission study, ten SPF cats were inoculated with saliva/blood collected from *Candidatus Mycoplasma turicensis* infected cats: five cats were inoculated subcutaneously (to mimic a cat bite), five cats received the inoculi oronasally (saliva) or orally (blood). The study comprised of five steps: I) saliva only; II) recipient cats immunosuppressed, saliva only; III) blood containing equivalent and IV) higher copy numbers than given previously in saliva; V) blood, s.c. (as positive control).

Table 1: Experimental setup

Group	Aim	N° of cats	Carrier	Route
Group A	I. Transmission by saliva	5	100 µl of saliva (120 copies)	subcutaneous
	II. Transmission by saliva, immunosuppression		2 x 200 µl of saliva (1x10 ³ copies)	subcutaneous
	III. Transmission by blood (twice in cat1)		10 µl of infectious blood (1x10 ³ copies)	subcutaneous
Group B	I. Transmission by saliva	5	2 ml of saliva (920 copies)	oronasal
	II. Transmission by saliva, immunosuppression		6 ml of saliva (7x10 ³ copies)	oronasal
	III. Transmission by blood		63 µl of infectious blood (8x10 ³ copies)	oral
	IV. Transmission by blood		500 µl of infectious blood (4x10 ⁵ copies)	oral
	V. Transmission by blood, positive control		50 µl of infectious blood (6x10 ⁶ copies)	subcutaneous

Results

Table 2: Outcome of the transmission study

Group A	I. Saliva sub-cutaneous	II. Saliva + Immunosuppression	III. Blood sub-cutaneous
Cat 1	●	●	●*
Cat 2	●	●	●
Cat 3	●	●	●
Cat 4	●	●	●
Cat 5	●	●	●

- PCR-negative
- PCR-positive, seropositive
- PCR-negative, seropositive

Using only saliva, neither oral nor subcutaneous inoculation led to infection in groups A and B independent of presence or absence of immunosuppression (I/II). When blood containing the same copy number was inoculated subcutaneously, infection was successful in four cats of group A (real-time TaqMan PCR positive), whereas cat 1 sero-converted after a second subcutaneous injection of 10 µl of blood (III). However, the oronasal group B stayed negative by PCR and serology after oral inoculation of blood (III/IV). When group B received the inoculi subcutaneously (positive control experiment), all five cats turned PCR-positive and seropositive (V).

Group B	I. Saliva oronasal	II. Saliva + Immunosuppression	III. Blood oral, 63 µl	IV. Blood oral, 500 µl	V. Blood sub-cutaneous
Cat 6	●	●	●	●	●
Cat 7	●	●	●	●	●
Cat 8	●	●	●	●	●
Cat 9	●	●	●	●	●
Cat 10	●	●	●	●	●

* twice in cat 1

Conclusion

Transmission by direct social contact via saliva seems highly unlikely. Oronasal exposure of up to 6 ml did not lead to transmission of infection. Transmission via aggressive interaction may occur if the recipient cat is exposed to a minimal volume of infectious blood (e.g., in the case of gingivitis/stomatitis of the aggressor cat).



Reference

- Neimark, H., Johansson, K.E., Rikihisa, Y., Tully, J.G., 2001. Proposal to transfer some members of the genera *Haemobartonella* and *Eperythrozoon* to the genus *Mycoplasma* with descriptions of *Candidatus Mycoplasma haemofelis*, *Candidatus Mycoplasma haemomuris*, *Candidatus Mycoplasma haemosuis* and *Candidatus Mycoplasma wenyoni*. Int. J. Syst. Evol. Microbiol. **51**, 891-899.
- Foley, J.E., Pedersen, N.C., 2001. *Candidatus Mycoplasma haemominutum*, a low-virulence eperythrozoon parasite of cats. Int. J. Syst. Evol. Microbiol. **51**, 815-817.
- Willi, B., Boretti, F.S., Cattori, V., Tasker, S., Meli, M.L., Reusch, C., Lutz, H., Hofmann-Lehmann, R., 2005. Identification, molecular characterization, and experimental transmission of a new hemoplasma isolate from a cat with hemolytic anemia in Switzerland. J. Clin. Microbiol. **43**, 2581-2585.
- Willi, B., Boretti, F.S., Tasker, S., Meli, M.L., Wengli, N., Reusch, C.E., Lutz, H., Hofmann-Lehmann, R., 2007. From *Haemobartonella* to hemoplasma: Molecular methods provide new insights. Vet. Microbiol. **125**, 197-209.
- Willi, B., Boretti, F.S., Meli, M.L., Bernasconi, M.V., Casati, S., Hegglin, D., Puorger, M., Neimark, H., Cattori, V., Wengli, N., Reusch, C.E., Lutz, H., Hofmann-Lehmann, R., 2007a. Real-time PCR investigation of potential vectors, reservoirs, and shedding patterns of feline hemotropic mycoplasmas. Appl. Environ. Microbiol. **73**, 3798-3802.